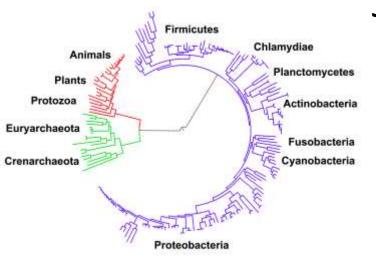




Plant Systematics and Plant/Pollinator Interactions

Jacob Landis



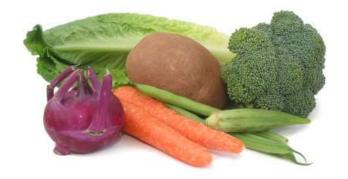


Why study plants

Important for food







Clothing







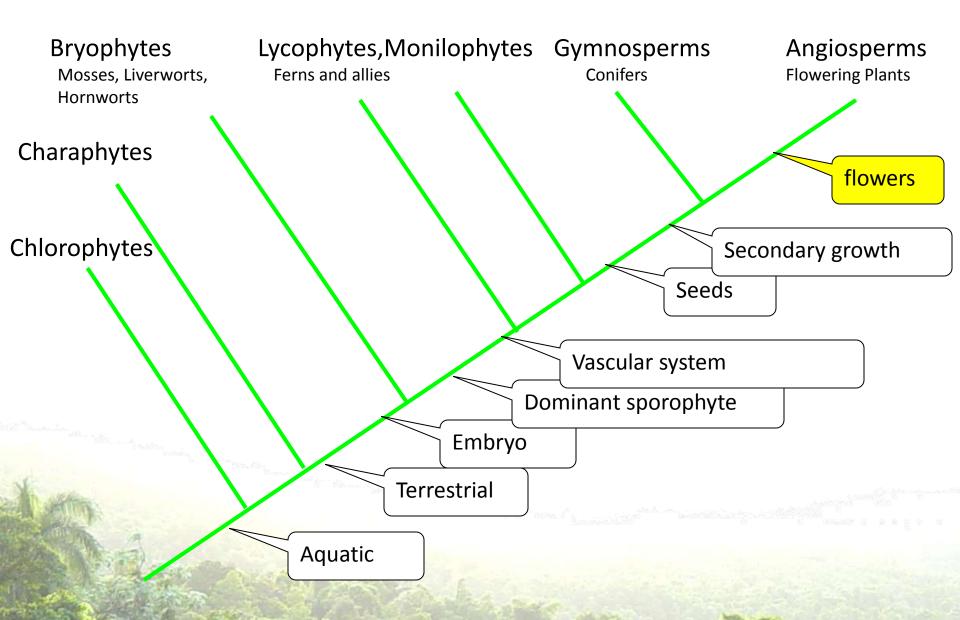
Drugs







BASIC PHYLOGENY OF PLANTS



Flower Diversity



Whorl name/number Function



1. Sepals 1. Protection

Whorl name/number

Function



1. Sepals



2. Pollinator attraction

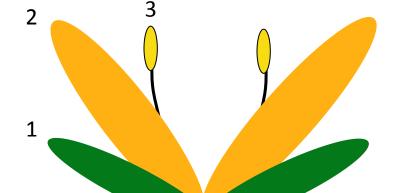
1. Protection

Whorl name/number

3. Stamens

2. Petals

1. Sepals



Function

3. Male gametes

2. Pollinator attraction

1. Protection

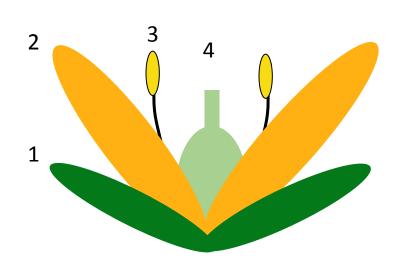
Whorl name/number

4. Carpels

3. Stamens

2. Petals

1. Sepals



Function

4. Female gametes

3. Male gametes

2. Pollinator attraction

1. Protection

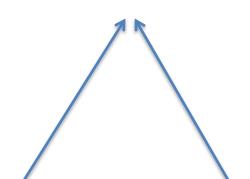
Research options

- Plant systematics
 ¬ relationships among plants
- Flower Evo-Devo

 how flowers develop, within or between species
- Plant/pollinator interactions
 why does the flower look the way it does to attract its pollinator, and how did it get that way

Goal





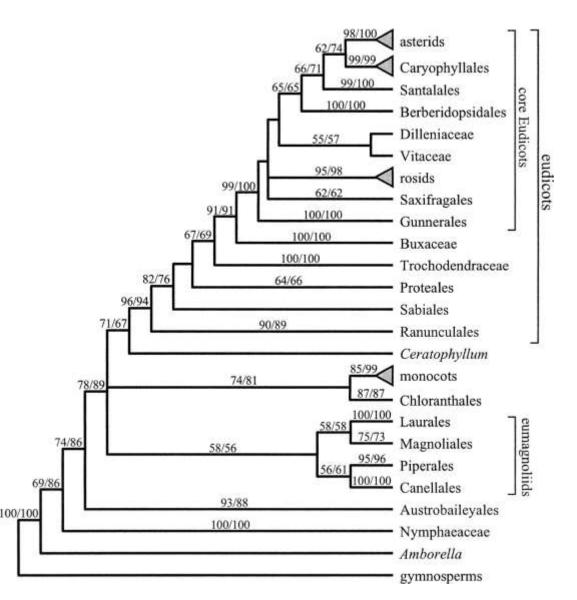




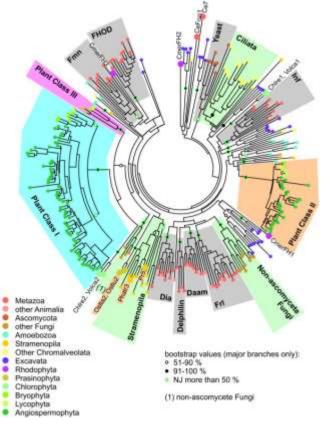


 Overarching goal is to link genetic changes to observable adaptations (specific pollinators)

Phylogenetics



- Shows
 relationships
 between
 organisms
- Important in understanding why plants are the way they are



Phylogenetics continued

- Conducted with either morphological or molecular data
 - historically morphology
 - predominately molecular now
- You will get hands on practice with both methods
 - compare which method is easy, more repeatable

POLLINATION MECHANISMS

Abiotic Pollination

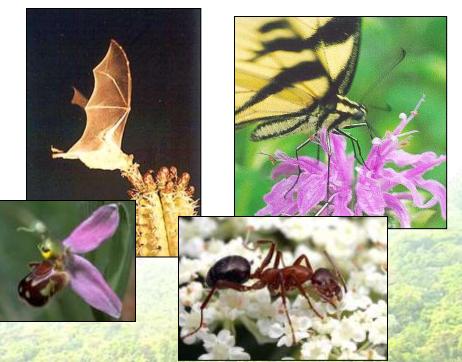
- Wind (anemophily)
- Water (hydrophily)

• Biotic Pollination

- Invertebrate
- Vertebrate







FLORAL ADAPTATIONS FOR POLLINATION

- Rewards
 - Pollen
 - Nectar
 - Oil and other substances
 - Protection
 - Breeding area
 - Sexual attraction (pheromones)

- Floral Cues
 - Color
 - Shape
 - Size
 - Scent

- <u>Efficiency</u>
 - Shape
 - Position
 - Distance
 - Pollen-stigma

TABLE 4.1 Floral pollination syndromes.

	Floral characteristics				
Pollinator	Color	Scent	Time of flowering	Corolla	Reward
Bee	Blue, yellow, purple	Fresh, strong	Day	Bilateral landing platform	Nectar and/or pollen
Butterfly	Bright; often red	Fresh, weak	Day	Landing platform; sometimes nectar spurs	Nectar only
Moth	White or pale	Sweet, strong	Night or dusk	Dissected; sometimes nectar spurs	Nectar only
Fly (reward)	Light	Faint	Day	Radial, shallow	Nectar and/or pollen
Fly (carrion)	Brownish, purplish	Rotten, strong	Day or night	Enclosed or open	None
Beetle	Often green or white	Various, strong	Day or night	Enclosed or open	Nectar and/or pollen
Bird	Bright; often red	None	Day	Tubular or pendant; ovary often inferior	Nectar only
Bat	Whitish	Musky, strong	Night	Showy flower or inflorescence	Nectar and/or pollen
Nonflying mammals	Dull-colored	Unscented to variously strong	Night	Robust, exserted styles and stamens	Copious nectar and/or pollen

BEE POLLINATION SYNDROME

- Two types:
 - Showy, <u>open</u>, <u>bowl-shaped</u>
 - Showy, <u>complicated</u>, <u>zygomorphic</u> (specialized)
- All be flowers tend to be <u>yellow, blue, or purple</u>
- UV nectar guides
- Sweetly scented
- Nectar and/or pollen





BUTTERFLY POLLINATION SYNDROME

Large and showy flowers with <u>landing platform</u>

Or large clumps of smaller flowers

Bright – <u>red, pink or lavender</u>

Long, narrow tubular flowers

• Fresh, weak scent

Nectar only





BIRD POLLINATION SYNDROME

- Large, showy flowers
- Red and tubular
- Exserted stamens/stigma
- Produce a lot of nectar
- Usually lack scent

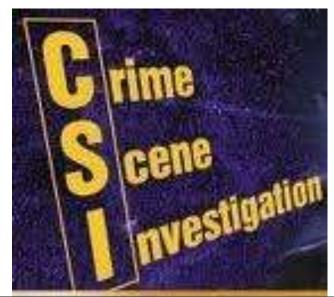


If someone asks you the following question:





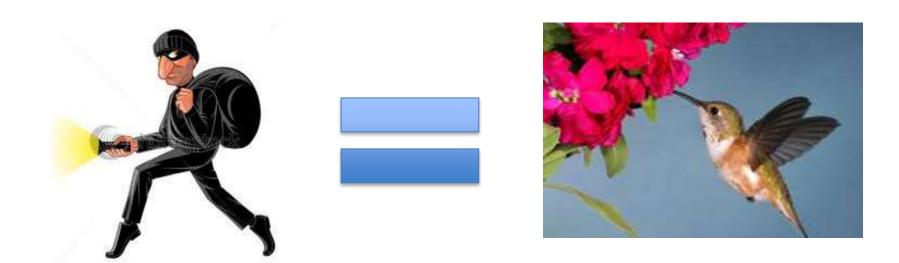
Science and CSI



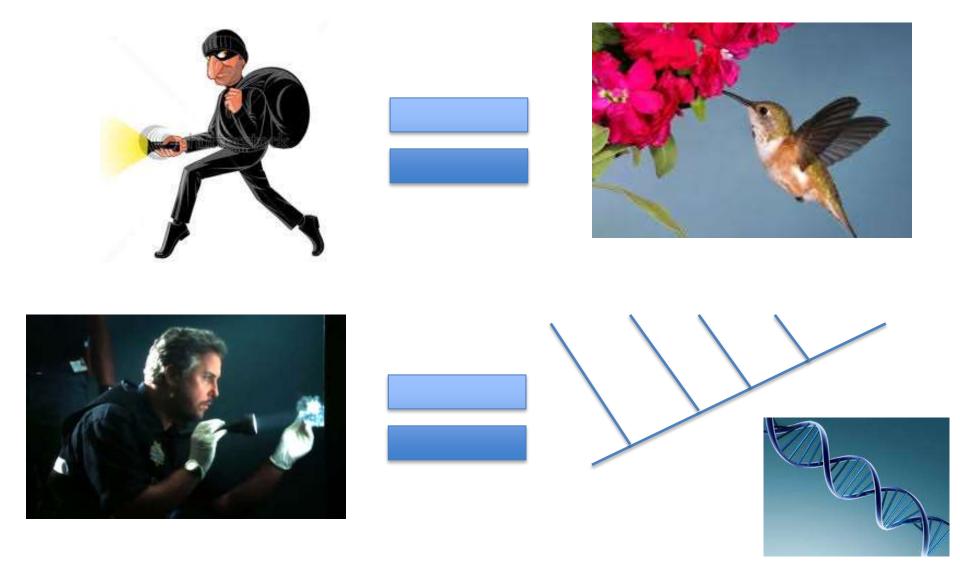




Comparison to CSI



Comparison to CSI



Goal for today

- Start comparing flowers and their morphology
- Extract DNA
- Run PCR
- Summary and prepare for Wednesday

The Plants

- 12 in total
- Different types of pollinators
 - Hummingbird
 - Bee
 - Butterfly



Hummingbird Pollinated



Maltese Cross



Petunia



Cardinal Climber

Bee



Snapdragon



Empress of India



California Poppy



Blue Daze

Butterfly



Pentas



Lantana



Vinca

Unknown



Blue Flax



Nicotiana

Thinking about flowers

PLANT MORPHOLOGY AND DNA EXTRACTION

1. Select four plants for morphological and genetic analysis. Record their names below.

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- 1	-	L
- 1		г
ı	_	•

ш,			
	Plant Number	Plant Name	Description of Plant
	1		
	2		
	3		
	4		

2. Observe and describe your four plants. As a group, identify floral characters you can use in order to group them by similarity. Which plants are most similar and why? Draw and describe below. Do you think the morphological relationships you describe also indicate genetic relatedness?

Hands on Techniques

 Any ideas why we would do the molecular techniques we will be learning today: extracting DNA and doing a PCR?

Hands on Techniques

 Any ideas why we would do the molecular techniques we will be learning today: extracting DNA and doing a PCR?

 Think about examples from popular TV shows, such as CSI?

CSI and who is the victim

http://www.youtube.com/watch?v=6iFDphW
 Xjw4



DNA extractions

- Label tubes
- Take hole punch of leaf material
- Extraction solution
- Cook at 95°C for 10 minutes
- Dilution solution





Follow protocol on hand outs to extract DNA from the four plants at your table

- Collection tubes
- Extraction buffer
- Dilution buffer

Now what do we do?

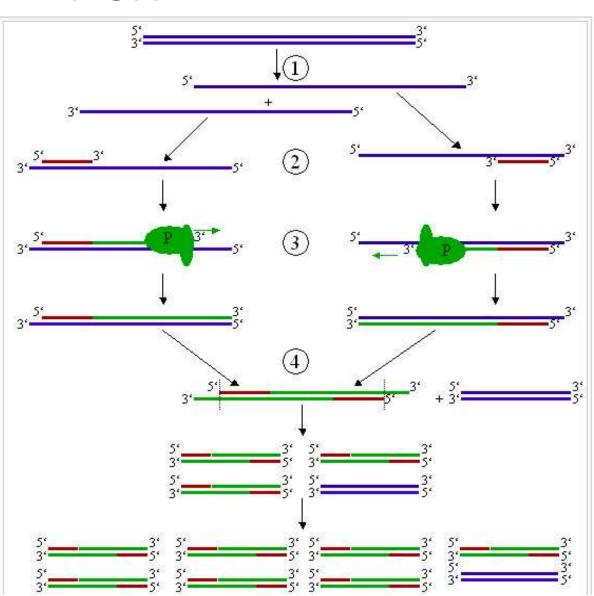
- We isolated DNA from the plant material into a solution form.
- Will this tell us what species the plant is, just like we found out the victim's identity?

Now what do we do?

- We isolated DNA from the plant material into a solution form.
- Will this tell us what species the plant is, just like we found out the victim's identity?
 - Not quite yet
- Additional step needed

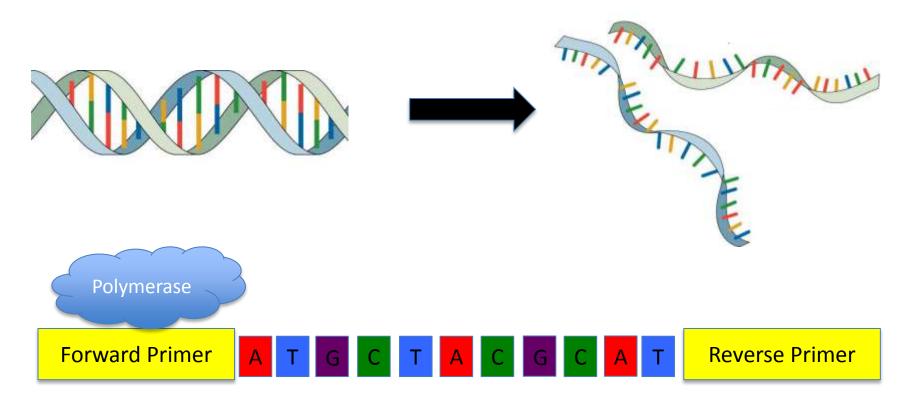
PCR

- PolymeraseChain Reaction
- Amplify small section of DNA, makes many many copies
- Will use common markers



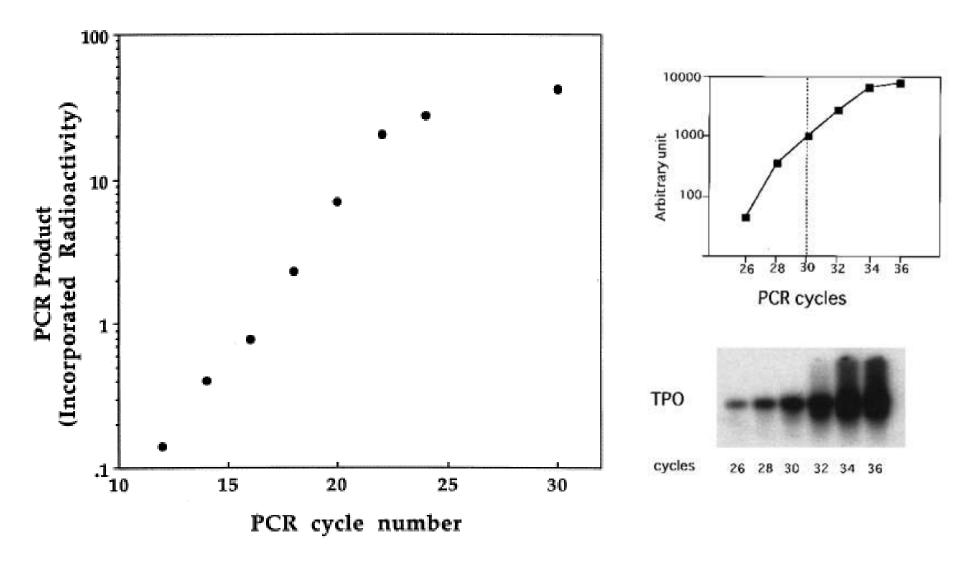
http://serc.carleton.edu/microbelife/research methods/genomics/pcr.html

How PCR works

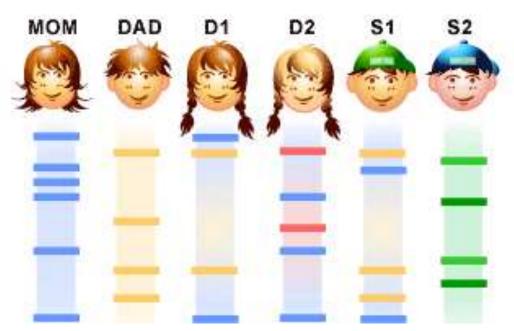


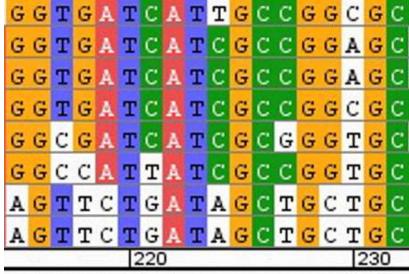
Melting
Annealing
Extending

Increase in product



Why do PCR?







Your turn to do a PCR

 Identify the following reagents at your student workstation, or collect from a common station:

Tube Label	Contents	V
PCR Mix REDExtract-N-Amp PCR Ready Mix		
	(Contains MgCl ₂ , dNTPs, Taq Polymerase)	
Water	Sterile water	
Forward	Forward primer	
Reverse	Reverse primer	
Positive	Positive control	
1-4	Your plant DNA samples from the previous procedure	

- 2. Vortex and centrifuge all reagents.
- Label six 0.2mL PCR tubes: 1-4, +, and -. Include your group identifier on each as well.
- 4. Prepare your PCR master mix in a clean 1.5mL microcentrifuge tube. You will perform 6 PCR reactions (4 leaf extraction DNA samples, positive control, and negative control). To ensure you have enough PCR master mix, you will prepare for 7 reactions. Be sure to change your tip between each reagent.

1 PCR Reaction	7 PCR Reactions	~
10μL REDExtract-N-Amp Ready Mix		
2μL of Forward Primer		
2μL of Reverse Primer		
2μL of Water		
4μL of Leaf Extract		
20μL Total Reaction		

Your turn to do a PCR

 Identify the following reagents at your student workstation, or collect from a common station:

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20μL Total Reaction		

Not in Master

Summary

- Extract DNA?
 - Why did we do this?

Summary

- Extract DNA?
 - Why did we do this?
- PCR
 - How does this work and why is it necessary?

Summary

- Extract DNA?
 - Why did we do this?
- PCR
 - How does this work and why is it necessary?
- Flowers
 - If you look at a flower, do you start getting ideas of what may pollinate it?





Goal for Wednesday

- Run PCR product on gels
- Talk about how we get sequence data
- Construct morphological phylogeny
- Construct molecular phylogeny
- Compare trees and the evolution of pollination syndromes

